PHYSIOLOGY

Effects of Nootropics on Electrical Activity in Rat Hippocampal CA1 Area

V. G. Motin, V. V. Yasnetsov, S. M. Kovalev, and I. N. Krylova

Translated from *Byulleten' Eksperimental'noi Biologii i Meditsiny*, Vol. 130, No. 9, pp. 252-254, September, 2000 Original article submitted October 6, 1997

Experiments on rat hippocampal slices showed that high concentrations of nooglutil and mexidol, but not piracetam, suppressed the ortho- and antidromic population responses in the CA1 area evoked by paired stimulation, the orthodromic responses being more sensitive to the drugs. The effects of both drugs were blocked by AP7, a specific antagonist of N-methyl-D-aspartate receptors. It is suggested that N-activation of methyl-D-aspartate receptors by high concentrations of nooglutil and mexidol reduced CA1 pyramidal cell responsiveness.

Key Words: nootropics; hippocampal slices; N-methyl-D-aspartate receptors

The mechanisms of nootropic action are now intensely studied *in vitro* on slice preparations of the hippocampus, which plays an important role in learning and memory [5,7-10]. It was found that nootropics change electrophysiological characteristics of rat and guinea pig hippocampus. They either facilitate long-term potentiation of synaptic responses in the CA1 and CA3 areas (pitacetam, oxiracetam, aniracetam, *etc.*) [11-13], or enhance short-term potentiation of synaptic responses in CA1 pyramidal cells induced by application of N-methyl-D-aspartate (NMDA) (piracetam, etimizol, nooglutil, ambocarb) [1]. This work was aimed at investigation of the effects of nooglutil and mexidol, original nootropic preparations [2-4] on evoked electrical activity in the CA1 area.

MATERIALS AND METHODS

The experiments were carried out on hippocampal slices $(350\text{-}400~\mu)$ from 38 Wistar male rats (150-200~g) [6]. The slices were perfused with a medium containing (mM): 126 NaCl, 3 KCl, 1.2 MgSO₄, 1.25 NaH,PO₄,

P. K. Anokhin Institute of Normal Physiology, Russian Academy of Medical Sciences; Hydrobioz Research-and-Production Center, Russian Ministry of Health, Moscow 2 CaCl₂, 26 NaHCO₃, and 10 glucose at a rate of 2 ml/min. The medium was saturated with carbogen (95% O₂ and 5% CO₂, pH 7.4) and heated to 35±0.5°C. Electrical activity was recorded after 1-h preconditioning in the perfusion medium. CA1 population responses were recorded with glass microelectrodes filled with 0.15 M NaCl. Ortho- and antidromic stimuli were applied to the Schaffer collaterals and alveus, respectively, through bipolar platinum electrodes. Single or paired square pulses (3-8 V, 0.1 msec) were used for stimulation. Population responses were recorded with an ATAK-350 analyzer. DL-2-amino-7-phosphonoheptanoic acid (AP7, Tocris Neuramine) was used as a specific NMDA receptor antagonist.

RESULTS

Piracetam (100 μ M-10 mM) did not affect the latency, amplitude, and configuration of population responses in rat hippocampal slices (n=10).

Nooglutil in concentrations from 10 μ to 5 mM (n=10) also had no effect, while in a higher concentration (10 mM, 50-min perfusion, n=5) it reversibly inhibited both the antidromic (Table 1) and, to a greater extent (p<0.002), orthodromic responses. The responses returned to the baseline after 1-h washout

V. G. Motin, V. V. Yasnetsov, et al.

Drug	Orthodromic responses		Antidromic responses	
	ı	II	1	П
Nooglutil, 10 mM	86±3	81±4	50±7	43±7
Mexidol, 5 mM	91±4	87±5	53±7	52±5

TABLE 1. Suppression of Ortho- and Antidromic Responses with Nooglutil and Mexidol (%, $M\pm m$)

(Fig. 1). No suppression was observed in the presence of NMDA receptor antagonist AP7 (20 μ M) introduced into the perfusion medium 15 min prior to nooglutil (10 mM, n=5) (Fig. 1).

Mexidol in a concentration of 5 mM (n=10) caused similar suppression (Fig. 2), while in lower concentrations (10 μ M-2.5 mM, n=10) it was ineffective. Orthodromic responses were also more susceptible to mexidol (p<0.001). Both ortho- and antidromic potentials returned to the baseline after 1-h washout (Fig. 2). Mexidol-induced suppression was also blocked by AP7 (n=5, Fig. 2).

Thus, both nooglutil and mexidol in high concentrations (but not piracetam) inhibited NMDA-dependent population responses in the CA1 hippocampal area induced by paired ortho- and antidromic stimulation. Synaptic transmission in the Schaffer collaterals-CA1 pyramidal cells system was very sensitive to nootropic-induced depression, because orthodromic potentials were less sensitive to the drugs.

The involvement of NMDA-receptors in this effect was not surprising. It was previously shown that specific NMDA receptors antagonist AP5 blocked the effects of oxiracetam on hippocampal CA1 field responses in rats [11]. Nootropics of different chemical structure enhance short-term potentiation of synaptic transmission in the CA1 region of hippocampal slices induced by NMDA [1].

Thus, nooglutil and mexidol in high concentrations inhibit synaptic transmission in the Schaffer collaterals-CA1 pyramidal cells system through activation of NMDA receptors.

REFERENCES

- I. I. Abramets, P. V. Andreev, and I. M. Samoilovich, *Eksp. Klin. Farm.*, No. 1, 15-17 (1995).
- T. A. Voronina, in: Current Problems of Experimental and Clinical Pharmacology [in Russian], Ed. V. E. Novikova, Smolensk (1994), pp. 28-30.
- 3. K. N. Dyumaev, T. A. Voronina, and L. D. Smirnov, *Antioxidants in Prophylaxis and Therapy of CNS Pathology* [in Russian], Moscow (1995).

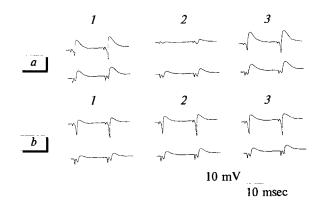


Fig. 1. Effects of nooglutil (10 mM, a) on orthodromic (upper curves) and antidromic (lower curves) responses in the rat hippocampal CA1 area induced by paired stimuli and their blockade with AP7 (20 μ M, b). 1) control; 2) exposure; 3) washout.

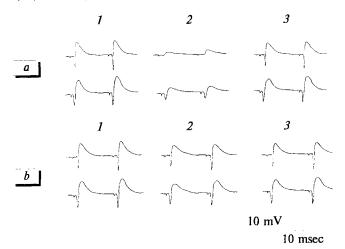


Fig. 2. Effects of mexidol (5 mM, a) on ortho- (upper curves) and antidromic (lower curves) responses in rat hippocampal CA1 area induced by paired stimuli and their blockade with AP7 (20 μ M, b). 1) control (a) or AP7 (b); a) exposure; 3) washout.

- S. B. Serednin and T. A. Voronina, Eksp. Klin. Farm., No. 1, 4-10 (1992).
- 5. A. N. Chepkova, N. V. Doreulee, R. U. Osrtrovskaya, et al., Byull. Eksp. Biol. Med., 110, No. 12, 602-604 (1990).
- V. V. Yasnetsov, V. M. Popov, Yu. P. Pal«tsev, et al., Ibid., 118, No. 12, 606-608 (1994).
- V. V. Yasnetsov and I. N. Krylova, *Uspekhi Fiziol. Nauk.*, No. 1, 97-116 (1997).
- 8. D. Ashton and L. Werbrouck, Brain Res., 541, 167-170 (1991).
- 9. V. K. Gribkoff, L. A. Bauman, and C. P. Van der Maelen, *Neuropharmacology*, **29**, 1001-1009 (1990).
- M. Marchi, E. Besana, and M. Raiteri, Eur. J. Pharmacol., 185, 247-249 (1990).
- A. M. Pugliese, R. Corradetti, L. Ballerini, and G. Pereu, *Br. J. Pharmacol.*, **99**, 189-192 (1990).
- M. Satoh, K. Ishihara, and Katsuki, Neurosci. Lett., 93, 189-192 (1988).
- 13. U. Staubli, M. Kessler, and G. Lynch, *Psychobiology*, **18**, 377-381 (1990).